AN ANESTHETIC ALKAMIDE AND FIXED OIL FROM
ACMELLA OLERACEA

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ABSTRACT: An anesthetic alkamide, (2E,4E,8Z,10E)-N-isobutyl-2,4,8,10-dodecatetraenamide (1) and fixed oil were isolated from the aerial part of the toothache plant, Acmella oleracea (L.) R.K. Jansen. Structure elucidation of 1 was achieved by analysis of 1H- and 13C-NMR experiments and comparison of the spectral data with published values. This is the first time to report 1 from A. oleracea (L.) R.K. Jansen. Additionally, fatty acids determination in lipid was obtained by GC.

Keywords: Acmella oleracea, local anesthetic, alkamide, (2E,4E,8Z,10E)-N-isobutyl-2,4,8,10-dodecatetraenamide

INTRODUCTION: The toothache plant or Paracress (Acmella oleracea (L.) R.K. Jansen, family Compositae) is renamed from the two toothache plants, Spilanthes acmella (L.) Murray and S. oleracea Jacquin. It is an annual herb that scattered around the world’s tropic. The plant has shown anti-inflammatory, antibacterial, antifungal, diuretic, sialagogic and larvicidal properties. The flower heads extract was used in India for treatment of gum diseases and dental caries. In Thailand, especially the Northern and North-eastern parts use A. oleracea (L.) R.K. Jansen as an indigenous vegetable and a spice for appetizers. This plant contains spilanthol (2) which was the principal pungent compound and acts as an anesthetic and analgesic. In this paper, phytochemical investigation of the aerial parts of A. oleracea (L.) R.K. Jansen is reported for isolation of (2E,4E,8Z,10E)-N-isobutyl-2,4,8,10-dodecatetraenamide (1), an anesthetic alkamide like spilanthol. Compound 1 has been isolated from A. ciliata, S. oppositifolia, Asarum forbesii, S. alba, Salmea scandens, Echinacea angustifolia, E. pallida, E. purpurea and Leucanthemum hosmariense. This is the first time to report the isolation of 1 from A. oleracea (L.) R.K. Jansen. Additionally, we also investigate fatty acids composition of fixed oil by GC analysis.

MATERIALS AND METHODS:

General experimental procedures
The TOF-MS data was obtained from a Q-TOF-2™ Micromass UK and NMR data were recorded from a Bruker Avance 400 FT-NMR spectrometer. Fatty acids composition was acquired by GC Shimadzu 2010.

Plant Materials
The aerial parts of Acmella oleracea (L.) R.K. Jansen, family Compositae were collected in Muang District, Chiang Mai Province, Thailand in September 2005. Authentication was achieved by comparison with the herbarium specimen at the Herbarium Section, Northern Research Center for Medicinal Plants, Faculty of Pharmacy, Chiang Mai University, Thailand. The herbarium specimen (No.5652) has been deposited in the Faculty of Pharmacy, Chiang Mai University, Thailand.

Extraction and isolation
A dried and powdered aerial parts of Acmella oleracea (L.) R.K. Jansen (1.5 kg) was macerated three times for a 3-day-period in 95% ethanol and filtered. The combined filtrate was evaporated under reduced pressure to give a dark green mass (12 gm). This mass (11 gm) was separated by vacuum column chromatography (VCC) of silica gel 60 (Merck, 70-230 mesh

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ASTM) and eluted with gradient technique of C_{6}H_{12}-EtOAc-CH_{3}OH to give 17 fractions. Fraction 1 eluted with hexane is yellow oil, was identified fatty acids composition by GC to give data as shown in Table 1. Fraction 6 eluted with 20-30% EtOAc in hexane, shows the most tingling effect on the tongue (onset = 2 seconds, duration = 20 minutes), was successively chromatographed on a Sephadex LH-20 column with IPA, silica gel column with C_{6}H_{12}-EtOAc-CH_{3}COCH_{3} (8:1:1) and with C_{6}H_{12}-EtOAc-CH_{3}COCH_{3} (12:1:1) to give a semipurify fraction. This fraction was then purified by preparative HPLC (Inertsil PREP-ODS, 10 x 250 mm, solvent system, MeOH-H_{2}O (75:25): flow rate, 5 ml/min; detector, UV 230 nm) to yield 1 (10 mg).

\[ (2E,4E,8Z,10E)-N\text{-}isobutyl-2,4,8,10-dodecatetraenamide \] (1) white solid, m.p. 63 °C, Micromass Q-Tof MS m/z (rel. int.): 248.1808 ((M +1) 5), 167.1154 (100).

$^1$H-NMR (400 MHz, CDCl_{3}) $\delta$ (ppm): 5.79 (1H, d, $J = 15$ Hz, H-2), 7.18 (1H, dd, $J = 15$, 10 Hz, H-3), 6.15 (1H, dd, $J = 15$, 10 Hz, H-4), 6.07 (1H, dt, $J = 15$, 7 Hz, H-5), 2.23-2.28 (2H, m, H-6), 2.23-2.28 (2H, m, H-7), 5.26 (1H, dt, $J = 10$, 7 Hz, H-8), 5.97 (1H, br t, $J = 10$ Hz, H-9), 6.29 (1H, br t, $J = 12.5$, H-10), 5.71 (1H, dq, $J = 15$, 7 Hz, H-11), 1.77 (3H, br d, $J = 6.8$ Hz, H-12), 3.15 (2H, t, $J = 7$ Hz, H-1), 1.80 (1H, m, H-2'), 0.93 (6H, d, $J = 6.4$ Hz, H-3', H-4')

RESULTS AND DISCUSSION: The fatty acids composition of fixed oil identified by GC was shown in Table 1.

An olefinic alkamide, \((2E,4E,8Z,10E)-N\text{-isobutyl}-2,4,8,10-dodecatetraenamide \) (1), is N-isobutylamide that exhibits strongly numbness and tingling effects like those of spilanthol. The tingling effect of these alkamides influence the sensory of trigeminal nerve. Furthermore, the structure of 1 presents the moieties as the 2,4-diunsaturated amide, 2E double bond and N-isobutylamide which had been studied to be important to this activity.\(^\text{[10]}\). The isolation and structure elucidation of 1 were succeeded by chromatographic and spectroscopic techniques together with comparison of the spectral data with published values. From the chemical shift and coupling patterns of protons in $^1$H-NMR data, we can assign the geometric isomer of 1 as \((2E,4E,8Z,10E)-N\text{-isobutylamide} \) but not as the other isomers because of these reasons; H-4 and H-5 were at downfield than those of the 4-cis. The $J_{4,5}$ was at 15 Hz instead of 11.5, as expected for a cis-double bond. The 10-trans geometry has more down field shift of H-10 and H-11 than those the 10-cis, and also from the $J_{10,11}$ of 15 Hz. Furthermore, chemical shifts and coupling constants for H-2, 3, 4, 5, 8, 9, 10 and 11 were similar to those reported for these protons in \((2E,4E,8Z,10E)-N\text{-isobutyl}-2,4,8,10-dodecatetraenamide \)\(^\text{[4,6,7,9]}\).

Interestingly, the fatty acids found in lipid presented an omega - 6 unsaturated fatty acid, linoleic acid (56.37%). This means that Acmella oleracea (L.) R.K. Jansen may be a rich source of essential fatty acid.

<table>
<thead>
<tr>
<th>Common name</th>
<th>IUPAC name</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Caprylic acid (C8:0)</td>
<td>Octanoic acid</td>
<td>0.14</td>
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<tr>
<td>Lauric acid (C12:0)</td>
<td>Dodecanoic acid</td>
<td>0.12</td>
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<tr>
<td>Myristic acid (C14:0)</td>
<td>Tetradecanoic acid</td>
<td>0.28</td>
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<tr>
<td>Palmitic acid (C16:0)</td>
<td>Hexadecanoic acid</td>
<td>25.84</td>
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<td>Palmitoleic acid (C16:1 n-7)</td>
<td>cis-9-Hexadecanoic acid</td>
<td>0.93</td>
</tr>
<tr>
<td>Stearic acid (C18:0)</td>
<td>Octadecanoic acid</td>
<td>4.54</td>
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<tr>
<td>Oleic acid (C18:1 n-9)</td>
<td>Octadecenoic acid</td>
<td>8.72</td>
</tr>
<tr>
<td>Linoleic acid (C18:2 n-6)</td>
<td>cis-9,12-Octadecadienoic acid</td>
<td>56.37</td>
</tr>
<tr>
<td>Linolenic acid (C18:3 n-3)</td>
<td>cis-9,12,15-Octadecatrienoic acid</td>
<td>0.72</td>
</tr>
<tr>
<td>Arachidic acid (C20:0)</td>
<td>Eicosanoic acid</td>
<td>0.66</td>
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<tr>
<td>Arachidonic acid (C20:4 n-6)</td>
<td>Eicosatetraenoic acid</td>
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<tr>
<td>Behenic acid (C22:0)</td>
<td>Docosanoic acid</td>
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<tr>
<td>Lignoceric acid (C24:0)</td>
<td>Tetracosanoic acid</td>
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<tr>
<td>Erucic acid (other 2C 22:1)</td>
<td>Cis-13-Docosanoic acid</td>
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</table>

![Chemical structure of 1](attachment://structure1.png)

![Chemical structure of 2](attachment://structure2.png)
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REFERENCES: